RESEARCH

Open Access



Retained introns in long RNA-seq reads are not reliably detected in sample-matched short reads

Julianne K. David^{1,2,3†}, Sean K. Maden^{1,2,4†}, Mary A. Wood^{1,5,6}, Reid F. Thompson^{1,2,7,8,9*} and Abhinav Nellore^{1,2,10*}

[†]Julianne K. David and Sean K. Maden contributed equally to this work.

*Correspondence: thompsre@ohsu.edu; anellore@gmail.com

 ² Department of Biomedical Engineering, Oregon Health & Science University, Portland, OR, USA
⁹ Department of Radiation Medicine, Oregon Health & Science University, Portland, OR, USA
Full list of author information is available at the end of the article

Abstract

Background: There is growing interest in retained introns in a variety of disease contexts including cancer and aging. Many software tools have been developed to detect retained introns from short RNA-seq reads, but reliable detection is complicated by overlapping genes and transcripts as well as the presence of unprocessed or partially processed RNAs.

Results: We compared introns detected by 8 tools using short RNA-seq reads with introns observed in long RNA-seq reads from the same biological specimens. We found significant disagreement among tools (Fleiss' $\kappa = 0.113$) such that 47.7% of all detected intron retentions were not called by more than one tool. We also observed poor performance of all tools, with none achieving an F1-score greater than 0.26, and qualitatively different behaviors between general-purpose alternative splicing detection tools and tools confined to retained intron detection.

Conclusions: Short-read tools detect intron retention with poor recall and precision, calling into question the completeness and validity of a large percentage of putatively retained introns called by commonly used methods.

Keywords: RNA-seq, Splicing, Intron retention

Background

During RNA transcription, multiple spliceosomes may act on the same transcript in parallel to remove segments of sequence called introns and splice together flanking exons [1]. Most splicing occurs stochastically [2] during transcription [3–5], although up to 20% of splicing may occur after transcription and polyadenylation [5, 6] (Additional file 1: Fig. S1). Introns are spliced by U2 and U12 spliceosomes [7], primarily in the nucleus [8], though studies suggest that cytoplasmic splicing may also occur [9–12].

Intron retention (IR) is a form of alternative splicing where an anticipated intron remains after transcript processing is complete. IR occurs in up to 80% of protein-coding genes in humans [13] and may affect gene expression regulation [14–20] as well as



© The Author(s) 2022. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http:// creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/public cdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.